Health Consultation

EXPOSURE INVESTIGATION

Investigation of Persistent Organic Pollutants in Reindeer on St. Lawrence Island

N. E. CAPE WHITE ALICE SITE (a/k/a N. E. CAPE SITE)

ST. LAWRENCE ISLAND, ALASKA

SEPTEMBER 21, 2001

U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES Public Health Service Agency for Toxic Substances and Disease Registry Division of Health Assessment and Consultation Atlanta, Georgia 30333

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In addition, consultations may recommend additional public health actions, such as conducting health surveillance activities to evaluate exposure or trends in adverse health outcomes; conducting biological indicators of exposure studies to assess exposure; and providing health education for health care providers and community members. This concludes the health consultation process for this site, unless additional information is obtained by ATSDR which, in the Agency's opinion, indicates a need to revise or append the conclusions previously issued.

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Prepared by:

Exposure Investigation and Consultation Branch Division of Health Assessment and Consultation Agency for Toxic Substances and Disease Registry

Summary of the Exposure Investigation

This exposure investigation examines reindeer, a significant source of food on St. Lawrence Island, for the presence of polychlorinated biphenyls (PCBs), polyaromatic hydrocarbons (PAHs), and organochlorine pesticides. This information will be provided to the local communities at Gambell and Savoonga, and will supplement information from the ongoing Reindeer Research Program at the University of Alaska, Fairbanks. In general, levels of all contaminants analyzed were low. We conclude that no health problems related to the group of chemicals analyzed in this exposure investigation would be expected in individuals consuming a diet containing large quantities of reindeer meat and fat.

Background

The Alaska Native community on St. Lawrence Island, Alaska, live near a formerly used defense site. This defense site, the Northeast Cape site, was used by the military for activities during and after World War II. This area has traditionally been used by the Siberian Yupik community for harvesting, hunting and fishing. Summer fishing camps are still located near the site.

Contaminants such as heavy metals have been found in the environment in previous work done by the Reindeer Research Program at the University of Alaska, Fairbanks. These contaminants have been detected in plants which are consumed by the free-ranging reindeer. These reindeer are a food supply for the local population which depends on a subsistence diet.

The question addressed by this exposure investigation is whether persistent organic pollutants might also be present in reindeer at levels which could result in health problems for the local population. This exposure investigation collected data to evaluate this pathway by analyzing reindeer blood (serum), meat (muscle), and fat (adipose tissue) from animals already being sampled by the Reindeer Research Program during the Year 2000 reindeer roundup. It was performed in conjunction with the U.S. Army Center for Health Promotion and Preventive Medicine (CHPPMS). This information will assist our response to the health concerns from the community.

Reindeer Overview

Reindeer (Rangifer tarandus) are semi-domesticated caribou. Although similar, there are fundamental differences in the behavior of reindeer and their wild cousins, caribou. Reindeer have been present in Eurasia for thousands of years. It is believed they have been domesticated there for at least 7000 years, which is longer than the horse (Edwards, 1994). In Eurasia reindeer are classified as either domesticated or wild, while in North America they are called reindeer if they are of the Eurasian domesticated variety, or caribou if they are of the wild variety. This domesticated factor makes them different from caribou in that they need to be tended on the range to keep them safe from predators, and may need to be driven to a better grazing area if theirs becomes sparse. They tend to be smaller than caribou, with shorter legs, and are a lighter color. They may have a life expectancy of 10-15 years, but mortality factors including disease and predation, not to mention unpredictable weather may cause them to die prematurely.

Reindeer eat lichen in winter and spring, and grass, birch and willow leaves, and mushrooms during the summer. They will always try to go to where the new greens are, which may be contrary to the direction the herder wants the herd to go.

Average reindeer weights (University of Alaska, Fairbanks-Reindeer Research Program).

Adults-	Female Average June Weight-	73.8 Kg	162.36 lbs.
	Female Average January Weight-	85.4 Kg	187.88 lbs.
	Male Average June Weight-	99.6 Kg	219.12 lbs.
	Male Average January Weight-	92.3 Kg	203.06 lbs.

Reindeer herding is an important cultural and economic part of life in many villages in Alaska. A concern expressed by some community members is that contaminants are being introduced into the food chain and compromising their health (Alaska Native Science Commission 1998). Current research programs, such as the Reindeer Research Program at the University of Alaska, Fairbanks, have analyzed heavy metals in reindeer, caribou, and native plants These heavy metals are readily absorbed by plants that are in turn eaten by ungulates, concentrating in liver, kidney, and muscle tissue (Gamburg and Scheuhammer, 1992).

Methods

This exposure investigation examined possible contamination of reindeer with PCBs, PAHs, and organochlorine pesticides. It was performed in conjunction with the U.S. Army Center for Health Promotion and Preventive Medicine (CHPPMS). Serum from 25 animals in last year's (2000) reindeer roundup was collected. Adipose tissue and edible meat (muscle) was collected from 8 animals which were already being sacrificed and tested under the University of Alaska Reindeer Research Program. Organ meat was not tested in this exposure investigation. Tissue and serum were shipped frozen. Analysis was performed by the U.S. Army Corps of Engineers Environmental Chemistry Branch Lab using standard EPA methodology for these substances (i.e. EPA 8270C, EPA 8081A, EPA 8082). Data validation was performed by Science Applications International Corporation, Oak Ridge, Tennessee.

Results

Table 1 (see Attachment 1) summarizes the data collected on PCBs, PAHs, and pesticides in reindeer serum, muscle, and fat. PCBs were not detected at levels above the 25 ug/L detection level. Organochlorine pesticides were also present only at levels slightly above the method detection limit and well below any health-based comparison values. Seven of the PAHs were present in levels which were accurately quantified.

Discussion

Reindeer are usually consumed from November through December, during the major herding time. Sometimes there is a hunt in July or August as well. Organ meat such as heart and liver may be eaten, however there is some disagreement over how commonly this happens. The hind and fore quarters, and spine meat, are regularly consumed. The fat is eaten and not trimmed off. Meat is generally boiled, but may also be fried, baked, broiled or dried.

In general, levels of all contaminants analyzed were low. (See attached table for specific values.) Only benzo (a) anthracene was present at a quantifiable concentration in muscle that was slightly above a health-based comparison value. Because a Minimal Risk Levels (ATSDR's health based comparison values) are currently not available for benzo (a) anthracene and many of the PAHs, EPA's oral reference doses or oral cancer slope factors were used as screening values for these chemicals.

Several other PAHs (benzo(b)fluoranthene, benzo(a)pyrene, indo(1,2,3-cd)pyrene, and dibenzo(a,h)anthracene) were found at levels above detection values, but too low to be accurately quantified. These four chemicals may, or may not, be at concentrations above their respective screening levels. It is important to note that these are *estimated* concentrations which were not considered accurately quantified.

Small amounts of benzo (a) anthracene were found in the muscle of 6 out of 8 reindeer slaughtered. The maximum concentration in muscle tissue was 22 ng/g (ppb). A risk calculation for an individual eating a large amount of reindeer meat and fat is included in Attachment 2. It is apparent from this risk calculation that consumption of reindeer meat and fat on St. Lawrence Island does not significantly increase an individuals risk of cancer.

Conclusions

1. Based on the analysis of PCB, PAH, and pesticide levels from the 2000 reindeer roundup, detectable health effects are not expected in individuals consuming reindeer muscle and fat on St. Lawrence Island.

Recommendations

- 1. Continue the usual local consumption patterns of reindeer meat and fat.
- 2. ATSDR will consider repeat analysis of these chemicals in a future reindeer harvest (2002 or 2003) to determine any change in contamination levels.
- 3. Consider analysis of organ meat (e.g. liver, kidney, heart) in any future sampling.

Prepared by: Robert Johnson Medial Officer Exposure Investigations and Consultations Branch Division of Health Assessment and Consultation

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References

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- 2. Agency for Toxic Substances and Disease Registry, Toxicologic Profile on Polychlorinated Biphenyls (Update), Atlanta, GA, November 2000.
- 3. Alaska Native Science Commission 1998. Traditional Knowledge and Radionuclides. University of Alaska Anchorage. 3211 Providence Drive, Anchorage, Alaska 99508.
- 4. Edwards, E.H., 1994, IN: The Encyclopedia of the Horse, Dorling Kindersley, pub., pp. 28-30.
- 5. Gamberg, M., and Scheuhammer, A.M. 1992. Cadmium in caribou and muskoxen from the Canadian Yukon and Northwest Territories, In: Sci. of the Total Environ. 143 (1194) 221-234.

Attachment 1

Table 1. Levels of PCBs, PAHs, and Pesticides in Reindeer Serum, Muscle, and Fat from the Savoonga Area.

Attachment 1

Table 1. Levels of PCBs, PAHs, and Pesticides in Reindeer Serum, Muscle, and Fat from the Savoonga Area.

Chemical	Serum			Muscle			Fat		
	detects	max.	m.d.l.	detects	max.	m.d.l.	detects		m.d.l.
	(#/total)	(ug/L)	(ug/L)	(#/total)	(ng/g)	(ng/g)	(#/total)	(ng/g)	(ng/g)
								<u> </u>	
Total PCB	0/25	n.d.	25	0/8	n.d.	3	0/8	n.d.	3
Naphthalene	0/12	n.d.	0.02	6/8	11 J	2.8	0/8	n.d.	2.8
2-Methylnaphthalene	0/12	n.d.	0.02	1/8	4.8 J	1.2	0/8	n.d.	1.2
Acenaphthylene	0/12	n.d.	0.02	1/8	3.9 J	0.8	0/8	n.d.	0.8
Acenaphthene	0/12	n.d.	0.02	4/8	14 J	0.8	0/8	n.d.	0.8
Fluorene	0/12	n.d.	0.02	4/8	19	1.7	0/8	n.d.	1.7
Phenanthrene	0/12	n.d.	0.02	8/8	67	1.2	0/8	n.d.	1.2
Anthracene	0/12	n.d.	0.02	6/8	40	1.5	1/8	3.7 ng/g J	1.5
Fluoranthene	0/12	n.d.	0.02	7/8	48	1.7	1/8	5.5 ng/g J	1.7
Pyrene	0/12	n.d.	0.02	7/8	40	1.2	1/8	5.5 ng/g J	1.2
Benzo (a) anthracene	0/12	n.d.	0.02	6/8	22	1.9	0/8	n.d.	1.9
Chrysene	0/12	n.d.	0.02	5/8	19	1.9	0/8	n.d.	1.9
Benzo (b) fluoranthene	0/12	n.d.	0.01	5/8	16 J	2.3	0/8	n.d.	2.3
Benzo (k) fluoranthene	0/12	n.d.	0.01	5/8	12 J	2.4	0/8	n.d.	2.4
Benzo (a) pyrene	0/12	n.d.	0.008	5/8	12 J	2.1	0/8	n.d.	2.1
Indeno (1,2,3-cd) pyrene	0/12	n.d.	0.01	4/8	8.7 J	2.7	0/8	n.d.	2.7
Dibenzo (a,h) anthracene	0/12	n.d.	0.009	3/8	6.8 J	2.8	0/8	n.d.	2.8
Benzo (g,h,l) perviene	0/12	n.d.	0.007	3/8	7.4 J	2.3	0/8	n.d.	2.3
Aldrin	0/25	n.d.	2.5 ug/L	0/8	n.d.	0.4	0/8	n.d.	0.4
Alpha BHC	1/25	2.0 J	2.5 ug/L	0/8	n.d.	0.5	0/8	n.d.	0.5
Beta BHC	1/25	2.3 J	2.5 ug/L	0/8	n.d.	0.8	0/8	n.d.	0.8
Delta BHC	2/25	2.0 J	2.5 ug/L	0/8	n.d.	0.4	0/8	n.d.	0.4
Gamma BHC (Lindane)	0/25	n.d.	2.5 ug/L	0/8	n.d.	0.4	0/8	n.d.	0.4
4,4'-DDD	0/25	n.d.	5.0 ug/L	0/8	n.d.	1	0/8	n.d.	1
4,4'-DDE	0/25	n.d.	5.0 ug/L	0/8	n.d.	0.4	0/8	n.d.	0.4
4,4'-DDT	3/25	5.0 J	5.0 ug/L	0/8	n.d.	0.4	0/8	n.d.	0.4
Dieldrin	0/25	n.d.	5.0 ug/L	0/8	n.d.	0.4	0/8	n.d.	0.4
Alpha endosulfan	0/25	n.d.	2.5 ug/L	0/8	n.d.	0.3	0/8	n.d.	0.3
Beta endosulfan	0/25	n.d.	5.0 ug/L	0/8	n.d.	0.4	0/8	n.d.	0.4
Endosulfan sulfate	0/25	n.d.	5.0 ug/L	1/8	0.87 J	0.8	0/8	n.d.	0.8
Endrin	0/25	n.d.	5.0 ug/L	4/8	3.0 J	0.8	0/8	n.d.	0.8
Endrin aldehyde	0/25	n.d.	5.0 ug/L	3/8	1.9 J	1	0/8	n.d.	1
Heptachlor	1/25	1.1 J	2.5 ug/L	4/8	0.56 J	0.5	0/8	n.d.	0.5
Heptachlor epoxide	0/25	n.d.	2.5 ug/L	0/8	n.d.	0.4	0/8	n.d.	0.4
Methoxychlor	0/25	n.d.	25 ug/L	0/8	n.d.	2	0/8	n.d.	2
Alpha chlordane	1/25	1.3 J	2.5 ug/L	1/8	1.0 J	0.4	0/8	n.d.	0.4
Gamma chlordane	0/25	n.d.	2.5 ug/L	0/8	n.d	0.4	0/8	n.d.	0.4
m.d.l. = method detection lin	mit								
n.d. = non-detect									
J = estimated concentration			<u> </u>	1			-		
- esumated concentration	·		1					·	

Attachment 2

Risk calculation for benzo (a) anthracene exposure:

Small amounts of benzo (a) anthracene were found in the muscle of 6 out of 8 reindeer slaughtered. The maximum concentration in muscle tissue was 22 ng/g (ppb). An individual who consumed 1 kg (2.2 lbs) of reindeer meat each day for 4 months of the year would, over a 70 year lifetime, have an excess cancer risk of 7.7×10^{-5} .

 $\frac{C \times CF \times IR \times EF \times ED}{BW \times AT} = \frac{22 \text{ ng/g } \times (10^{-6} \text{ mg/ng } \times 10^{3} \text{g/kg}) \times 1 \text{ kg/d } \times 4 \text{ mo/12 mo } \times 70 \text{ yr}}{70 \text{ kg } \times 70 \text{ yr}} = 1.05 \times 10^{-4} \text{ mg/kg-d}}$

Dosage x CSFo = $1.05 \times 10^4 \text{ mg/kg-day} \times 0.73 (\text{mg/kg-day})^1 = 7.7 \times 10^{-5} \text{ excess cancer risk}$

C = concentration in meat/fat CF = conversion factors of ng to mg and g to kg IR = ingestion rate EF = exposure frequency ED = exposure duration BW = body weight AT = averaging time CSFo = oral cancer slope factor*

*Note that this is a provisional cancer slope factor and may reflect much uncertainty in predicting risk.

Sample Numbers for Serum Samples Only	Sample Numbers for Serum Samples w/associated fat & muscle
1 (serum)	samples with the same sample number
2 (serum)	22 (serum, fat & muscle)
3 (serum)	23 (serum, fat & muscle)
6 (serum)	24 (serum, fat & muscle)
8 (serum)	25 (serum, fat & muscle)
9 (serum)	26 (serum, fat & muscle)
11 (serum)	27 (serum, fat & muscle)
12 (serum)	29 (serum, fat & muscle)
13 (serum)	30 (serum, fat & muscle)
14 (serum)	Total number of samples = 24 (serum, fat & muscel)
16 (serum)	
17 (serum)	
18 (serum)	
19 (serum)	
20 (serum)]
21 (serum)	
31 (serum)	

Total Samples = 17 serum

Total Number of Serum samples only 17

Total Number of Serum samples with corresponding fat & muscle samples 8

Total number of fat samples 8

Total number of muscle samples 8 Total number of samples 41

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	y papers in the appropriate place?
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